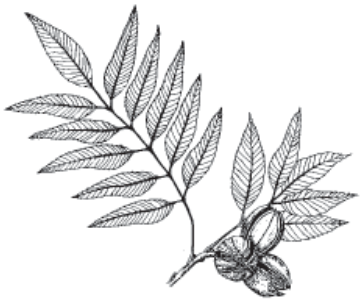
OKLAHOMA PECAN GROWERS ASSOCIATION

Volume LI, No. 3 *Michael Smith, Editor* *July, August, September 2010*

In This Issue



Oklahoma Pecan Growers' Association is published 4 times per year and is a benefit of being an association member. Contact the Oklahoma Pecan Growers' Association c/o Horticulture & Landscape Architecture, Oklahoma State University, 358 Agriculture Hall, Stillwater, OK 74078-6027 for further information.

President's Corner

I would like to thank all of those that made the 2010 OPGA summer conference such a success. In particular, I would commend the Savage Family for the field day at their farm Hauani Creek. Not only did they cart around 190 people to see their entire impressive operation but they feed us all as well. Basil, Francis, their children, and grand children went all out for our benefit. Thanks!

The arrangements for the rest of the show were made by Charles Rohla, Janice Landgraf and Randy Bryant. The Sunday evening meal was donated, without charge, by Farm Credit. The new convention facility at Ardmore was perfect for our needs. This facility had plenty of air conditioned space to exhibit any sort of pecan equipment indoors, as well as room for classrooms and the banquet. The banquet food was as good as we have ever had. The food show winner auction raised about $3500 for pecan research at OSU. The Mount family once again furnished the food show awards. Andrea Mount organized the judging. Thanks also to Dr. Mike Smith for arranging the educational program. Finally, thanks to all of the speakers that were involved in the Monday sessions.

Congratulations to Mark Sandmann on being selected grower of the year. The Herman Hinrichs award went to the Noble Foundation for all that they have done for us.

We have made arrangements to have the 2011 OPGA conference in Bartlesville at the new Hilton Gardens Inn. The dates are June 26 and 27 with the field day on Tuesday June 28, 2011. Individual reservations can be made at www.bartlesville.hgi.com.

Janice Landgraf has served as treasurer of the OPGA for the last 6 years. We need someone to step-up to take her place. She has done a great job. Thanks Janice! If you or anyone you know wish to be treasurer contact me or Charles Rohla the incoming President.

Bob Knight
918-321-6011

The Changing Face of Extension
Eric T. Stafne

I have been a member of the OSU cooperative extension service for almost five years. Now, depending on your perspective five years is either a very long time or just a drop in the bucket. For me it has flown by, but the way that I approach my extension programming is changing. Let's take a look back at the way extension programming was done in the past – one-on-one field visits, publication of fact sheets, answering phone calls, holding field days, and a lot of driving around the state. To a certain extent some of those are still being done;

however, with new technology and less funding, the methods of information dissemination are changing. Five years ago I was doing the traditional extension stuff, but today I don't have the luxury of driving around the state whenever someone needs me to look at their trees. I especially cannot do many one-on-one visits or if I do they need to be on a day when I do multiple visits in the same area. The new way of performing extension programs is to have as many people as possible listen, read, or see your material at one time. Therefore, my Pecan Management Short Course with 25 students makes much more economic sense than driving 2 hours to visit one grower. In other states such visits are banned – no more one-on-one visits. In Oklahoma we haven't yet reached that level, but it may not be far off.

As a way of increasing the access to extension material I am relying more on the internet. Just for example, I write a blog now for the grape industry. The grape industry has a much different demographic than the pecan industry and a blog may not work for the pecan industry. When I attended the recent OPGA conference in Ardmore I did not see many fresh, new faces that will be steering the industry for the next 30 or 40 years. This worries me. What is the succession plan for the Oklahoma pecan industry? What is the succession plan for research on pecans in Oklahoma? Dr. Mulder gave a presentation on the new Pecan ipmPIPE project. It sure was impressive how much work had been done on that project and the amount of information available, but in my opinion the jury is still out on how much impact it will have on pecan growers in Oklahoma. Maybe I am underestimating the computer savvy-ness of our growers, but I still get many requests from pecan growers who have no computer access (or desire to use one).

Luckily, extension is highly valued in Oklahoma. There is a lot of support within OSU for cooperative extension and we will continue to serve the industry the best we possibly can in a multitude of different ways. But as budgets decrease year after year you might see a little less of our faces in person and a little more of them on your computer screen.

Early Crop Overview for Southern Oklahoma
Charles Rohla, Noble Foundation

The 2010 pecan crop in southern Oklahoma appears to be substantially larger than we have had in several years. The past two years the Noble Foundation orchard at Burneyville has suffered from freeze events. The improved cultivars did not have a crop either year and the native crop has been light. The season started off with an extremely heavy flower set on the improved cultivars and above average crop on the natives. We had a small drop during pollination, however on the orchard as a whole crop set remained heavy. Pecan nut casebearer flight was light and we elected to hold off spraying for a few days to allow some damage to occur to lighten the crop. We started spraying the orchard 3 days after we found the first entry. Upon further evaluation of the crop we were still seeing 70-75% of terminals with fruit. We did observe a later capture of moths just prior to spraying, approximately 14 days after firth moth capture. Several growers called wondering if they should spray or not. Most were not finding eggs or entries. Some growers elected to go ahead and spray while other did not. I have visited with several growers that did not spray and damage ranges from little to one orchard that has reported over 80% damage from pecan nut casebearer. This definitely shows the importance of monitoring throughout casebearer season especially if you do not spray.

Fig. 1. Ovule (kernel) one-half expanded

With the heavy crop several growers are starting to call wondering if the heavy crop will affect next year's crop. Most likely if you have a heavy crop this year the crop will be lighter next year, unless you lighten the crop through fruit thinning during July and August. Some have thought that since this is an 'off' year that we will have another good crop next year. We can pray for one, but the likelihood of having two good crops are slim, unless you are managing your trees to reduce stress associated with heavy crops.

Fruit thinning can be very beneficial in improved cultivar orchards. Not all cultivars respond to fruit thinning. Native trees are typically too large to remove enough nuts and the smaller nuts do not weigh enough to be shaken from the tree. Research from 1934 done on 'Moore' pecan trees showed that it took at least 8 to 10 functional

leaves to adequately fill one nut. Therefore, if you have large clusters or larger nut cultivars more leaves are required to fill the nuts. Removing a portion of the crop on trees with a heavy crop has the potential to increase nut size and nut quality (percent kernel and grade) and increase return bloom the following year. Through fruit thinning, several managers have been able to maintain a more consistent crop year to year.

The time to thin depends on development of the kernel. To determine the development of nuts is done by cutting the nuts to expose the ovule (Fig. 1). Thinning should be done when the ovule or kernel is at least 50% expanded and before the kernel is in dough stage. Thinning is achieved using a tree shaker equipped with donut pads. The donut pads are important to protect the trees from trunk damage. Greasing between the rubber flaps and the donut pads will also help provide trunk damage.

To determine if you need to fruit thin and to determine how much fruit to remove, you need to first determine the percent of terminals with fruit. To calculate the percentage of terminals with fruit, count 100 terminals in mid-canopy and record how many have at least one fruit. If your cultivars have large nut sizes (less than 50 nuts per pound) the optimum load is 45-50% terminals with fruit. If your cultivars are moderate size nuts (50-70 nuts per pound) the optimum load is 50-60% terminals with fruit. If your cultivars are smaller (greater than 70 nuts per pound) the optimum load is 60-70% terminals with fruit. The use of an observer on the ground to watch a predetermined area and evaluate the percentage of fruiting terminals is very beneficial. Do not look at the number of nuts on the ground, focus on what remains in the tree.

Department of Agriculture
Farm Service Agency

7 CFR Parts 760 and 783
Commodity Credit Corporation
7 CFR Part 1416
RIN 0560-AH96

Tree Assistance Program
AGENCY: Farm Service Agency and Commodity Credit Corporation, USDA.
ACTION: Final rule.

SUMMARY: This rule implements specific requirements for the Tree Assistance Program (TAP) authorized by the Food, Conservation, and Energy Act of 2008 (the 2008 Farm Bill). TAP provides disaster assistance to eligible orchardists and nursery tree growers to replant or rehabilitate trees, bushes, and vines that were lost due to natural disaster. Orchardists and nursery tree growers who commercially raise trees, bushes, and vines for which there were mortality losses in excess of 15 percent, after adjustment for normal mortality, are eligible for TAP payments. Eligible losses must have occurred between January 1, 2008, and September 30, 2011. This rule specifies how the TAP payments are calculated and when producers may apply for benefits. This rule also removes regulations for prior tree disaster assistance programs.

Contact your local FSA for additional information.

Horticulture Pecan Research Endowment
Michael Smith
OSU Horticulture & Landscape Architecture

As of this writing contributions total $56,950, and the Oklahoma Pecan Growers' Association has pledged $20,000. This is your opportunity to make a difference in the future of Oklahoma pecans and O.S.U. research and education. Creation of an Endowed Professorship will ensure that pecan research at O.S.U. continues indefinitely. Remember, our goal is to reach $250,000 to get matching funds from the T. Boone Pickens gift and the State of Oklahoma. Checks should be made out to the **O.S.U. Foundation** and mailed to **Michael Smith, Department of Horticulture and Landscape Architecture, 358 Agricultural Hall, Oklahoma State University, Stillwater, OK 74078.** Contributions to the Endowment are tax deductable.

Below is a list of those contributing to the Endowment.

<u>2008</u>
Paul and Maxine Haydon
Bert and Elizabeth Blumer
J.D. and Dwayne Scott
G.F. Parsons
Edward L. Boyd, Jr.
John Barnes
Henry Bellmon
Alvin and Debra Stein
Michael and Carole Smith
Virginia Merritt Autry
Tim Montz
Bag-A-Nut, LLC

<u>2009</u>
Joe Ihle
Diane Couch
Terry D. Powell
George Carlson
Dean McCraw
Carole and Max Matheson
Bill Ault
Glenn Taylor
Williams Companies
Paul and Maxine Haydon

<u>2010</u>
Michael and Carole Smith
Irvin R. (Bud) Blakley
Bob Hightower

Why the Early Fungicide Applications?

Damon Smith
OSU Entomology & Plant Pathology

If you happened to attend the 2010 Oklahoma Pecan Growers Association (OPGA) Meetings held in Ardmore, Oklahoma a couple of weeks ago then you probably got a chance to see my presentation on managing pecan scab with fungicides. I reported the results of the 2009 fungicide trial, which we conducted at the Cimarron Valley Research Station located in Perkins, OK. If you were unable to attend the meeting the methods and results of those trials can be found below.

While presenting the data, I mentioned that the first application in each of the fungicide programs was applied as an early pre-pollination spray without the assistance of the Oklahoma Mesonet Pecan Scab Advisor. That is to say that the first spray was a phenological-based spray, which was applied prior to catkin opening. All subsequent sprays were then applied based on the Oklahoma Mesonet Pecan Scab Advisor. I have received many questions inquiring about the theory behind the timing of the first application of fungicide. The theory is actually based on concepts that were introduced by two pioneering plant disease epidemiologists. J.C. Zadocks and R.D. Schein published a text titled "Epidemiology and Plant Disease Management" in 1979. In the text they introduced three ways to slow plant disease epidemics. According to Zadocks and Schein, one way to slow an epidemic is to eliminate or reduce the initial inoculum (spore) level. This will translate to a reduced level of initial disease. With pecan scab epidemics, levels of initial inoculum can be reduced by performing winter pruning, using good sanitation practices, or using very early fungicide applications. The second way to slow a disease epidemic is to slow the rate of disease increase. This is typically accomplished by using pecan varieties that are resistant to scab and/or by using periodic fungicide sprays. Finally, Zadocks and Schein indicated that a plant disease epidemic could be reduced by shortening the time of exposure of the crop to the pathogen. In row crops this is easily accomplished by adjusting planting dates. In a perennial crop like pecans, however, this strategy is not easily accomplished.

With pecan disease management we can feasibly rely upon methods one and two. In figure 1, I have demonstrated that using combinations of management strategies to reduce both the initial inoculum level and the rate of increase can affect a disease epidemic. The black line indicates an epidemic when no management strategies were used. The gray line illustrates what can happen if the initial inoculum level and the rate of disease increase are influenced by good disease management strategies. Note that disease still occurs when management strategies are used, but the amount and rate of disease increase is substantially reduced.

So how does all of this relate to the early pre-pollination fungicide applications we applied in our fungicide evaluations? By applying those early applications, I theorize that we have managed to reduce initial spore load of the scab fungus. In turn this reduces the amount of initial disease. By combining this with the subsequent fungicide applications applied according to the Oklahoma Mesonet Scab Advisor, the rate of disease increase can also be reduced. We have applied treatments again this season to verify this theory. Hopefully the results will be consistent with those of 2009.

2009 Oklahoma State University Pecan Fungicide Evaluation

The fungicide trial was established at Cimarron Valley Research Station, Perkins OK. Rootstocks ('Colby' seedlings) were planted on an upland site with a Konawa fine sandy loam soil in 1994. Established rootstocks were grafted using 'Maramec' scion in 1996 and 1997. The experimental design was a randomized complete block with

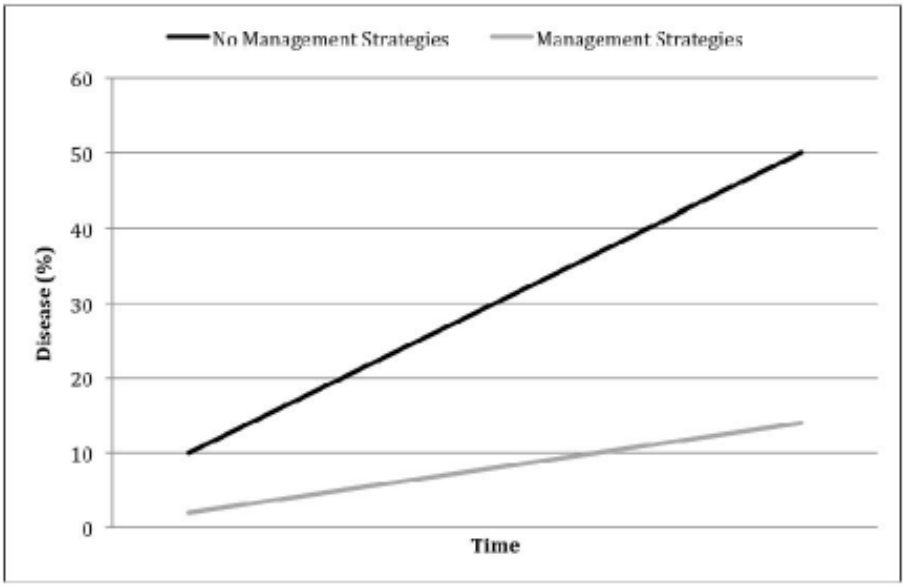


Figure 1. The effect of disease levels over time when using no scab management strategies (black line) vs. using scab management strategies (grey line). Note the difference in initial disease level (lower starting point) and reduced rate of disease increase (decreased slope of the line) for the treatment where management strategies were used (grey line).

four replicates. Plots consisted of two trees with at least one border tree between adjacent plots on all sides. Trees were spaced 35 ft apart with a between-row spacing of 35 ft. Recommended maintenance practices were followed throughout the growing season, including crop-load thinning. Fungicides were applied with a tractor mounted Nut Hustler air blast sprayer, calibrated to deliver 104 GPA.

First fungicide applications were applied on 4 May (pre-pollination). Subsequently, treatments were applied using the Oklahoma Mesonet Pecan Scab Advisor (http://agweather.mesonet.org/index.php/data/section/hort) which was modified so that a scab hour was calculated based on a temperature threshold of 65°F and 85% relative humidity. Ratings of leaf incidence (percent of leaves with symptoms of pecan scab), leaf severity (average percent of leaf area with symptoms of pecan scab), fruit incidence (percent of clusters with symptoms of pecan scab) and fruit severity (average percent of cluster area with symptoms of pecan scab) were taken as an average of 8 branch terminals on each tree on 19 May, 11 Jun, 1 Jul, 7 Aug, and 4 Sep. Pecans were harvested on 6 Nov, dried to 6.5% moisture and yield data recorded on 9 Nov. Area under the disease progress curve (AUDPC) were assessed for five ratings of the percentage of leaves or fruit with symptoms of scab, averaged over 8 branch terminal ratings per tree. Leaf disease incidence and severity data were subjected to the area under the disease progress curve (AUDPC) transformation to account for season-long ratings. These data and final fruit incidence, severity, and yield data were analyzed using ANOVA (a = 0.05). Treatment means were compared by Fisher's test of protected least significant difference (a = 0.05).

Table 1. Results comparing four fungicide programs and a non-treated control for managing pecan scab in Oklahoma.

Treatment and rate/A	Timing[z]	Leaf Incidence AUDPC (%-days)[y]	Leaf Severity AUDPC (%-days)[x]	Final Fruit Incidence (%)[w]	Final Fruit Severity (%)[v]	Yield (lb/a)[u]
Non-treated control	N/A	3991.8 A	647.6	76.9 A	10.6 A	270.2
LATE STROBILURIN						
Folicur 3.6F 8.0 oz	1					
Enable 2F 8.0 oz	2,4					
Topsin 4.5FL 20.0 oz	3					
Headline 2.09EC 7.0 oz	5	2429.6 B	402.1	18.8 B	2.4 B	206.2
QUILT						
Quilt 1.66SC 27.5 oz	1,3,5					
Topsin 4.5FL 20.0 oz	2,4	1371.1 BC	215.0	4.7 B	0.8 B	397.0
STRATEGO						
Stratego 2.08EC 10.0 oz	1,3,5					
Topsin 4.5FL 20.0 oz	2,4	1109.0 BC	240.1	8.8 B	2.9 B	404.2
EARLY STROBILURIN						
Headline 2.09 EC 7.0 oz	1					
Topsin 4.5FL 20.0 oz	2,4					
Abound 2.08EC 12.3 oz	3					
Enable 2F 8.0 oz	5	806.4 C	350.7	2.4 B	2.0 B	304.2

[z]Application timing of each fungicide within each 5-spray sequence. N/A indicates not applicable.
[y]Means followed by the same letter are not significantly different according to Fisher's test of protected least significant difference where: LSD=1208; R^2=0.78; CV=41; P-value <0.01.
[x]Area under the disease progress curve (AUDPC) ratings per two-tree plot: R^2=0.64; CV=55; P-value = 0.08.
[w]Means followed by the same letter are not significantly different according to Fisher's test of protected least significant difference where: LSD=21; R^2=0.88; CV=62; P-value<0.01.
[v]Final fruit severity based on ratings (4 Sep) of average % per fruit cluster. Means followed by the same letter are not significantly different according to Fisher's test of protected least significant difference where: LSD=3.5; R^2=0.81; CV=62; P-value<0.01.
[u]Average yield (moisture = 6.5%) measured on 9 Nov: R^2=0.46; CV=41; P-value=0.21.

Results- and Discussion

Weather throughout most of the season was wet with below-normal temperatures. Abnormally hot weather was documented for a period of time in June. However, this weather did not persist. Highest levels of leaf disease incidence were recorded in non-treated check plots (Table 1). All fungicide programs resulted in significantly lower levels of leaf disease incidence compared to non-treated control plots (see table). Lowest levels of leaf disease incidence were recorded in plots treated according to the EARLY STROBILURIN program and were not significantly different from plots treated according to QUILT and STRATEGO programs. Plots treated according to the LATE STROBILURIN program had the highest levels of leaf disease incidence among plots where fungicide was applied. However, leaf disease incidence was not significantly different from QUILT and STRATEGO programs. Highest levels of leaf disease severity were observed in the non-treated control plots. No significant differences in leaf severity were noted among fungicide treatments and non-treated control plots. Fruit disease incidence and severity were highest in non-treated control plots. All fungicide programs resulted in lower levels of fruit disease incidence and severity compared to the non-treated control plots. However, no significant differences in fruit disease were noted among fungicide programs. No significant differences in yield were recorded among treatments.

Leaf Analysis Standards for Pecan – Revised 2010

Michael Smith
OSU Horticulture & Landscape Architecture

July is the month to take leaf samples in Oklahoma. Samples should consist of 100 middle leaflet pairs collected from the middle leaf on current season's growth. The sample should be collected from several trees that are representative of the orchard. Leaflets should then be rinsed in clean tap water and laid out to dry on newspaper or cardboard. If leaflets are not washed concentrations of foliar applied nutrients, such as zinc, will higher than what has been actually been absorbed by the leaf. If samples are soaked too long (longer than 1 minute) potassium will be leached from the leaf thus its values will be unreliable.

Samples should be taken to your local County Extension Educator where they will be transferred to a laboratory for analysis and recommendations. You have two choices for analysis. One is the Noble Foundation which has handled the samples for several years. However, they have changed labs and now charge $28/sample. Samples can also be sent to OSU's Soil, Water, and Forage Analytical Laboratory where the same analysis costs $20.

Based on recently completed research nitrogen, phosphorus, potassium, nickel and copper leaf analysis standards and recommendations have changed. The new recommendations are listed below. We now differentiate between native pecan trees and cultivars, so it is important to indicate what the sample is. All cultivars are treated similarly, so the exact cultivar is unimportant.

This research and the revised standards and recommendations have been made possible by funding from the Oklahoma Pecan Growers' Association, Samuel Roberts Noble Foundation, Fertilizer Check-off Program, Savage Equipment Company, and Oklahoma Agricultural Experiment Station. Without this support this research would not be possible.

Nitrogen

Native trees:

<2.3%	Low, double present N application rate. If the orchard did not receive N last year apply 150 lbs/acre N.
2.3 – 2.5%	Normal. Continue present N application rate.
2.5 – 2.7%	Normal. Nitrogen application can be reduced without affecting yield or nut quality. Decrease application rate by 20%.
2.7 – 3.0%	Normal. Nitrogen application can be reduced without affecting yield or nut quality. Decrease application rate by 50%.
>3.0%	Above normal. Normally, N concentrations of 3% or greater usually are associated with tree drought stress or excessive N application. Excess N application can exacerbate drought stress. Withhold all N for one year.

Cultivar trees:

<2.3%	Very low, double present N application rate. If the orchard did not receive N last year apply 150 lbs/acre N.
2.3 – 2.4%	Low. Increase the present N rate by 30%. If the orchard did not receive N last year, apply 125 lbs/acre N.
2.4 – 2.7%	Normal. Continue the present N application rate. If none was applied, then none will be needed.
2.7 – 3.0%	Normal. Nitrogen application can be reduced without affecting yield or nut quality. Decrease application rate by 30%.
>3.0%	Above normal. Normally, N concentrations of 3% or greater usually are associated with tree drought stress or excessive N application. Excess N application can exacerbate drought stress. Decrease application rate by 50%.

Nitrogen can be rapidly lost when soils are flooded or water saturated. Sites that are subject to occasional flooding may benefit from a split N application with 60% applied before budbreak and 40% applied about mid May. Sites with sandy soil may also benefit from a split N application. A single application before budbreak is adequate for other sites.

Phosphorus

Native trees:

<0.12%	Low. Apply 100 lbs/acre P_2O_5.
≥0.12%	Normal. None needed.

Cultivar trees:

<0.14%	Low. Apply 100 lbs/acre P_2O_5.
≥0.14%	Normal. None needed.

Potassium

Native trees:

<0.85%	Low. Apply 100 lbs/acre K_2O.
≥0.85%	Normal. None needed.

Cultivar trees:

<1.0%	Low. Apply 100 lbs/acre K_2O.
≥1.0%	Normal. None needed.

Calcium

<0.70%	Low. Apply lime based on soil test information.
≥0.70%	Normal, none needed.

Magnesium

<0.30%	Low. Soil test for pH. If low, use dolomitic limestone to adjust soil pH. Otherwise apply $MgSO_4$ at the manufacturer's recommendation.
≥0.30%	Normal, none needed.

Manganese

<100 ppm	Apply three foliar applications of $MnSO_4$ (32% Mn) at 6 lbs/acre of material beginning as the first leaf unfurls during budbreak, and then with the 1st generation pecan nut casebearer spray (late May to early June), and the 2nd generation pecan nut casebearer spray (late June to early July). Mn can be tank mixed with zinc and most pesticides. Other commercial Mn products may be used following the manufacturer's recommendations.
≥100 ppm	Normal, none needed.

Zinc

<60 ppm	**Mature trees** – apply 3 foliar applications of $ZnSO_4$ (36% Zn) at 6 lbs/acre of material beginning as the first leaf unfurls during budbreak, and then with the 1st generation pecan nut casebearer spray (late May to early June), and the 2nd generation pecan nut casebearer spray (late June to early July). Other Zn products may be used following the manufacturer's recommendations. If no Zn has been applied as a foliar application and the leaf Zn concentration is ≥50 ppm, then none will be needed. **Young trees** – apply foliar applications of ZnSO4 (36% Zn) at 6 lbs/acre of material beginning as the first leaf unfurls during budbreak, and then at two week intervals until shoot growth ceases. Other Zn products may be used following the manufacturer's recommendations.
≥60 ppm	Normal. Follow current Zn program.

Iron

<50 ppm	Iron deficiency can be induced by cool, wet environmental conditions in the Spring that inhibit Fe translocation. Improved environmental conditions will permit translocation and the shortage will be eliminated. Corrections of Fe deficiency will normally not be needed unless the shortage persists for two years or the deficiency is acute.
≥50 ppm	Normal.

Copper

<6 ppm	Low. Apply $CuSO_4$ or chelated Cu at the manufacturer's recommendations.
6 – 20 ppm	Normal, none needed.
>20 ppm	Excess.

Nickel

<2.5 ppm	The lower threshold for Ni sufficiency has not been firmly established. Trees with less than 2.5 ppm may benefit from Ni application. Nickel Plus is currently the only product available. Follow the manufacturer's recommendations.
≥2.5 ppm	Normal, none needed.

Classified ads may be placed in the Newsletter for free by OPGA members. Send your ad to Mike Smith at mike.smith@okstate.edu and it will appear in the next newsletter and subsequent Newsletters until notification to remove the advertisement.

ELECTRO-SPRAY

Pecan, fruit and vegetable sprayer. 3-pt mount, PTO powered, ten nozzles, 110 gal. tank, electric shut-off valve. Can spray heights up to 40-50 feet. Always stored in shed, $1,295.00. Call (918) 682-9448.

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Native Pecan 7 ft. $10.00
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wallsfarm@sbcglobal.net
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FOR SALE – FMC 4300 Diesel shaker $13,000. Call Tim Montz 940-733-0956.

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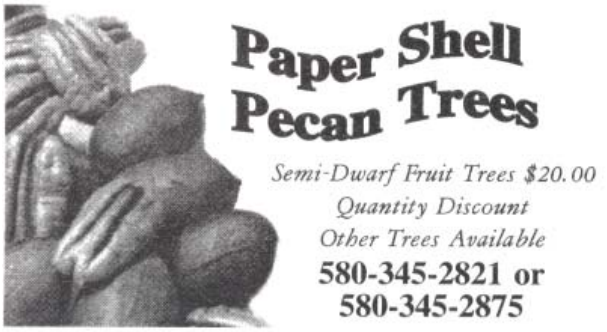

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Oklahoma Pecan Growers' Association
c/o Horticulture & Landscape Architecture
Oklahoma State University
358 Agricultural Hall
Stillwater, OK 74078-6027

Return Service Requested

Membership Application

We invite you to become a member of the Oklahoma Pecan Growers' Association. Membership includes the *OPGA Newsletter*, *Pecan South* and *Pecan Grower*. Make your checks payable to OPGA and mail to:

Oklahoma Pecan Growers' Association
Janice Landgraf, Treasurer
RR 1 Box 148
Madill, OK 73446
okpecan@trinex.net (580) 795-7644

Name _____

Street Address _____

City, State, Zip _____

Phone ( ) _____ email: _____

❑ Renew ❑ New Member

Grower Member $50.00
Industry Member $125.00
Extension/Research/Student $40.00